**Institutional Animal Treatment and Cadaver Usage at Quincy University**

**Mission Statement**

Quincy University, as a Franciscan Catholic institution, supports the advancement of knowledge while avoiding the causation of undue suffering to animals. All animal research at Quincy University is under the purview of the committee members of the IRB.

This document is designed to improve the quality of research proposals that utilize animals, make the animal care and use regulatory process easier for investigators/teachers, and most importantly, ensure the welfare of the animals.

**Guidelines and Policies**

Euthanasia

1. Invertebrates. The submission of a protocol is not generally necessary. Needless harm to animals is to be avoided. For example, pinning should not be used to hold live insects in place, when another method of restraint could be used. When needed, animals should be anesthetized by standard methods, such as CO2 narcosis or cold. Euthanasia by freezing or standard chemical methods (ethyl acetate, ethanol, acetone) should be used.
2. Vertebrates. The submission of a protocol is required for all but observational field studies.
   1. Fish  
      Euthanasia by freezing is acceptable.
   2. Amphibians  
      Euthanasia by pithing, Tricaine methane sulphonate (MS 222) immersion, barbiturate overdose (ip), or freezing is acceptable.
   3. Reptiles  
      Euthanasia by pithing, barbiturate overdose (ip), or freezing is acceptable.
   4. Birds and Mammals  
      For euthanasia of rodents and rabbits, methods of choice include carbon dioxide asphyxia, barbiturate overdose (iv or ip) and anesthetic overdose. Other acceptable methods in anesthetized animals only include decapitation, cervical dislocation and exsanguination. For euthanasia of birds, methods of choice include barbiturate overdose (iv or ip) and anesthetic overdose. Other acceptable methods include carbon dioxide asphyxia, and, in anesthetized animals only, decapitation, cervical dislocation and exsanguination.

It is assumed that other species (e.g. farm animals, primates) will be used rarely if at all at QU. In such cases, guidance should be sought in advance from the HASC committee, and a reputable source, such as the web site above, should be consulted.

*Specialized references:*

Guidelines to the use of wild birds in research. Gaunt, A.S. and Oring, eds. 2010. The Ornithological Council. https://birdnet.org/info-for-ornithologists/guidelines-to-the-use-of-wild-birds-in-research/guidelines-english-3rd-edition-2010/

Guide for the Care and Use of Agricultural Animals in Agricultural Research and Teaching, published by the Federation of American Societies of Food and Animal Science. https://www.fass.org

Federal Compliance

USDA Regulations require that each research facility register with the Secretary of Agriculture if the facility has USDA-regulated species on its premises. USDA-regulated species include all warm-blooded vertebrates except mice of genus *Mus* and rats of genus *Rattus* bred for research purposes and birds. Quincy University does not maintain any USDA-regulated species on its premises. Hence, it is not required to register.

Institutions receiving support from the Public Health Service for activities involving animals must provide an Assurance of Compliance (Assurance) with the PHS Policy. As QU does not currently enjoy any grant support from said agency, it is not bound to provide such an assurance.

Surviving surgeries

In accordance with good scientific practice and standards set forth in the Public Health Service Guide for the Care and use of Laboratory Animals (https://www.nap.edu/catalog/12910/guide-for-the-care-and-use-of-laboratory-animals-eighth) and the Federal Animal Welfare Act, aseptic surgical procedures and appropriate anesthesia must be used for all vertebrate surviving surgeries. Anesthesia of mammals prior to invasive procedures should be done in accordance with methods described on the University of Iowa web site: <https://animal.research.uiowa.edu/iacuc-guidelines-anesthesia>. Aseptic technique is not required if the animal has already received a lethal level of anesthetic.

Field work

Appropriate permits are likely to be required by law and should be obtained prior to collecting, trapping, or other field research on vertebrates other than unobtrusive observation. State or federal agencies should be consulted for their specific requirements.

Housing

**Animal health and husbandry should be discussed with a faculty advisor to apply state and federal guidelines.**

**Cadaver Policies**

Cadavers come from individuals who have bequeathed their bodies to science, hoping to make a contribution to medical education and/or scientific research and thereby improve the quality of human life and future health care for others. In appreciation, all cadaver donors shall be treated with respect.

Students enrolled in the cadaver anatomy course must adhere to the following guidelines and rules. In addition, students will carefully follow any supplementary policies provided by the instructor, whether in oral or written form. Any abuse of cadavers, unprofessional conduct, security breaches, or violation of policies described below will lead to an F in the course and possibly other penalties.

**Admittance to the Cadaver Anatomy Laboratory.**  Only the biology faculty and students enrolled in the course are allowed to be in the laboratory. Under absolutely no circumstances are unauthorized visitors permitted. Special permission for visitors must be sought from the instructor. Unauthorized individuals will be asked to leave the lab.

**Security.**  The cadaver will be housed in a laboratory dedicated solely to the purpose of the cadaver anatomy course. This space will be behind two lockable doors. These doors are to remain locked at all times except when enrolled students and faculty are present. Custodians and physical plant personnel are discouraged from entering the cadaver laboratory. All clean-up will be performed by the students.

**Photography**. Unauthorized photography in the cadaver laboratory is not permitted.

**Food and Beverages.** Absolutely no eating or drinking is permitted in the cadaver laboratory at any time.

**Laboratory Attire.** Protective clothing must be worn in the cadaver laboratory: lab coats and gloves. Shorts, open-toed shoes or sandals are not permitted.

**Contact Lenses**. Wearing of contact lenses is not recommended. Students that cannot wear eye glasses shall wear goggles at all times when in the lab. Everyone shall wear protective eye gear when using bone saws or chisels.

**Pregnancy.** Women who are pregnant (or who may potentially become pregnant) during the time of this course should consult their personal physician regarding risks associated with exposure to chemicals (isopropyl alcohol, ethanol, phenol, glycerin and formaldehyde).

**Anatomical/Cadaver Materials.**

Anatomical materials may NOT be removed from the laboratory. Any extraneous parts are to be frozen for later cremation. Students will NOT cut or deface any portion of the cadaver body unless so directed by the instructor.

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